

# Reviews

# The Practical Benefits of Pharmacokinetics in the Use of Antineoplastic Agents

C. Erlichman\*, R. C. Donehower, and B. A. Chabner

Clinical Pharmacology Branch, Clinical Oncology Program, Division of Cancer Treatment, National Cancer Institute, Bethesda, Maryland 20205, USA

Summary. Clinical pharmacokinetics has clearly had an important impact on the use of antineoplastic agents, but this influence has primarily been the result of comprehensive analyses conducted at the time of initial clinical trial. Such studies have often determined the dose, schedule, and route of administration, and have provided general guidelines for dose adjustment in patients with organ dysfunction. On the other hand, routine pharmacokinetic monitoring, while a highly effective adjunct to drug therapy in clinical specialties other than cancer, has not yet had an important effect on clinical oncology, with the obvious exception of high-dose methotrexate therapy. A number of potentially important applications of routine monitoring are pointed out in this paper, and will certainly be examined in the future as a means of dealing with pharmacokinetic variability. However, major impediments in this effort are posed by complexity of antineoplastic pharmacology and the lack of suitably sensitive, specific, and rapid assays for routine clinical use. Radioimmunoassays and competitive binding methods offer considerable hope in this effort, but the widespread application of these methods has not yet been realized in clinical practice.

### Introduction

An understanding of pharmacokinetics has become essential to the effective use of many classes of drugs, including antibiotics, cardiovascular agents, and anticonvulsants. In these areas, routine drug level monitoring has become commonplace, and has been of clear benefit in solving specific problems regarding drug delivery, optimization of therapeutic effect, and minimization of tox-

icity. It is not difficult to convince the average practitioner of the benefits of monitoring such agents as phenytoin or procainamide; it is well established that standard doses of phenytoin (300 mg per day) produce suboptimal plasma concentrations in over 50% of patients, and toxic levels in 16% [43]. Similarly, the original schedules of procainamide administration (every 6 h) have been adjusted to shorter time intervals (every 3 h) to accommodate the brief plasma half-life of this agent [61]. Similar useful applications of pharmacokinetic approaches have been infrequent in the field of antineoplastic therapy, principally because of the extreme chemical reactivity of many of these agents, their activity at low plasma levels, and the lack of suitably sensitive, rapid, and specific assay methods. However, this situation is rapidly changing thanks to major new advances in methods as well as clinical interest in basing therapy on pharmacologic considerations such as drug concentration and exposure duration. It will be the purpose of this review to discuss the impact of comprehensive pharmacokinetic studies and routine drug monitoring on current antineoplastic therapy.

Comprehensive pharmacokinetic investigations of new antineoplastic agents have been conducted for many years, and in many instances have provided useful information. In general, however, these studies were severely limited by the insensitivity of available assay methods, and often had to rely on the use of radioactive drugs. Such studies led to incomplete or mistaken conclusions, for several reasons: (1) Impure radiochemicals were used; (2) the radiolabel, such as labile [3H] atoms, often dissociated from the compound before administration or during transit through the body; and (3) total radioactivity was taken indiscriminately to be a reflection of parent compound, as in the case of studies of melphalan [72] and of methotrexate [37]. The introduction of highly sensitive and specific methods, such as radioimmunoassay, competitive protein binding, high-pressure liquid chromatography, and gas chromatography-mass spectrometry, has greatly altered this situation in the past 5 years, and has al-

<sup>\*</sup> Fellow of the Medical Research Council of Canada Reprint requests should be addressed to: C. Erlichman, Building 10, Room 6N119, National Cancer Institute, Bethesda, Maryland 20205, USA

Table 1. Assay methods available for selected antineoplastic agents

Agent	Assay method	References
Methotrexate	CPBA <sup>a</sup> , RIA <sup>b</sup>	2, 5, 12, 36, 46, 47, 54, 60
5-Fluorouracil	HPLC°, GC	15, 22, 23, 48
Cytosine arabinoside	HPLC, RIA	44, 58
6-Thioguanine	HPLC	49
6-Mercaptopurine	HPLC	49
Adriamycin	HPLC	39, 57
Bleomycin	RIA	13, 30
Melphalan	HPLC	20
Chlorambucil	HPLC	45
Cyclophosphamide	GC-MSd	34
Hexamethylmelamine	GC	29

- <sup>a</sup> Competitive protein-binding assay
- <sup>b</sup> Radioimmunoassay
- <sup>c</sup> High-pressure liquid chromatography
- d Gas chromatography-mass spectrometry

lowed precise measurement of parent compound and metabolites for many useful anticancer drugs (Table 1). These assays have been applied in two basic types of studies — comprehensive analyses of pharmacokinetics of new and established drugs, and routine clinical monitoring to aid adjustment of doses and clinical decision making in individual patients.

For the purposes of this discussion, comprehensive pharmacokinetic studies are considered to be detailed studies of drug disappearance in plasma, while routine drug monitoring differs in that a single time point is chosen as a valid reflection of the drug's pharmacokinetic behavior in that patient. For routine monitoring to be useful, comprehensive pharmacokinetic studies must have been completed for that drug.

# I. Comprehensive Pharmacokinetic Studies in Antineoplastic Therapy

The primary goals of comprehensive pharmacokinetic studies are to define the absorption, distribution, elimination, and metabolism of a drug, with the ultimate aim of achieving an optimal clinical schedule and dose [31]. The four primary elements of pharmacokinetics set the foundation for a rational approach to drug administration, and, if performed at the time of initial drug trials in man, may profoundly influence subsequent drug usage. The importance of each of these elements will be considered in the following discussion.

Drug absorption studies are critical to the choice of route of administration. Variable or incomplete oral absorption leads to unpredictable or ineffective therapeutic activity. Melphalan and hexamethylmelamine are examples of antineoplastic drugs for which variability in plasma levels after oral administration has been well documented. Both agents are usually administered in fixed doses. Alberts et al. [3] and Tattersall et al. [72] have reported wide interindividual variability in melphalan absorption. Melphalan excretion in feces after oral administration was documented to be between 20% and 50% of the administered dose. The ratio of concentration by time  $(C \times T)$  for oral versus intravenous administration of melphalan in three patients ranged from 0.3 to 1.0, while one patient showed no plasma melphalan after an oral dose. Even greater variability in plasma levels of hexamethylmelamine has been demonstrated by D'Incalci and co-workers, who found 100-fold differences in the C×T product in their study of 11 patients [28]. In this case, the variability may be attributable to either erratic absorption or variable first-pass metabolism of the parent compound by the liver. These data imply that the standard dosage regimens often used in the treatment of multiple myeloma and ovarian cancer with these agents may result in inadequate drug bioavailability, and support an increased use of drug level monitoring with dose modification to assure adequate therapeutic exposure.

Knowledge of drug distribution is also an important element in therapeutic decision making for specific tumors and clinical situations. Nitrosoureas are the preferred agents for treatment of central nervous system malignancies because of their high lipid solubility and ability to cross the blood-brain barrier [27]. However, studies of the distribution of non-lipid-soluble drugs such as methotrexate have also yielded unexpected and useful information [9]. The drug levels (above 1  $\mu$ *M*) that can be achieved in the cerebrospinal fluid during high-dose systemic infusion have allowed the successful use of this form of treatment for lymphoma metastatic to the central nervous system [64] and led to studies of its use as CNS prophylaxis in the treatment of childhood acute leukemia. Significant levels were also found in third spaces, such as peritoneal and pleural effusions [8]. The subsequent slow efflux of methotrexate from these compartments resulted in prolongation of the half-life of the drug in the systemic circulation, leading to prolonged exposure of normal tissues to low but cytotoxic concentrations of drug, with resultant unexpected toxicity. Recognition that the peritoneal space may selectively retain non-lipid-soluble agents such as methotrexate and 5-fluorouracil has resulted in the novel approach of using peritoneal dialysis as a means of delivering high concentrations of chemotherapy to intraperitoneal tumor, with limited exposure of systemic tissues [41, 65].

Knowledge of drug elimination, which most commonly occurs through renal excretion or hepatic metabolic degradation and biliary excretion, enables dose modification to be made in the face of organ dysfunction, or allows for continuation of maximal doses if this dysfunc-

tion is known not to influence pharmacokinetics. For example, studies of adriamycin elimination have revealed predominant elimination by metabolism in the liver, with subsequent biliary excretion. Toxicity was found to be increased in patients with hepatic compromise who received standard doses of the drug [7]. These findings led to the recommendation of 50% dose reduction for bilirubin levels above 1.5 mg per 100 ml, and 75% for bilirubin above 3 mg per 100 ml. Unfortunately, the available assays for adriamycin are not sufficiently simple or reliable to allow routine patient monitoring, and attempts to correlate blood levels with hepatic function or toxicity have been unsuccessful. Clearly there is a need for routine monitoring capability in the clinical use of adriamycin.

An additional example of the value of pharmacokinetic information is provided by recent studies of the combination of 5-fluorouracil (5-FU) and thymidine. These studies were initiated largely on the basis of the preclinical work of Martin and colleagues, who demonstrated agmented 5-fluorouracil incorporation into tumor RNA and enhanced antitumor activity in several murine systems [50, 51]. Clinical trials of this combination have not answered the question of its therapeutic value, but significant host toxicity has been observed [42, 74, 77]. When 15 g thymidine was administered prior to intravenous injection of 5-fluorouracil, the half-life of 5-fluorouracil was increased from less than 30 min to 6 h [77]. Increases renal clearance of intact drug and decreased [14C]-CO<sub>2</sub> production from labeled drug suggest that this enhanced potency is the result of competition of 5-fluorouracil and thymine (derived from thymidine) for pyrimidine-catabolic enzymes. Additional mechanisms of interaction at the cellular level are not ruled out by these studies, but a plausible explanation for increased clinical toxicity has been provided by pharmacokinetic analysis.

An appreciation of the requirement of certain antitumor agents for activation by hepatic microsomes may influence drug usage. Prednisone [71], hexamethylmelamine [62], cyclophosphamide [25], and imidazole carboxamide (DTIC) [17] are all agents that require metabolic activation by liver microsomes. Intra-arterial administration of any of these drugs is unlikely to confer any therapeutic advantage. The conclusion reached in a recent report of the use of intra-arterial DTIC in an adjuvant protocol for treatment of malignant melanoma was that this therapy does not provide any additional benefit [6]. Although other factors may have contributed to this negative result, the pharmacological considerations would have suggested that no advantage could possibly result from this route of administration as against intravenous therapy.

In spite of the rapid accumulation of pharmacokinetic data about chemotherapeutic agents, a caveat must be recognized because of the nature of the antineoplastic

process. Prediction of therapeutic or toxic effects on the basis of plasma pharmacokinetics presupposes that changes in plasma drug levels reflect changes within target tissues. However, the neoplastic process is heterogeneous with respect to differentiation, metabolic activity, clonogenic potential, and degree of vascularity. This heterogeneity may complicate the correlation of therapeutic effect with classic pharmacokinetic parameters such as plasma drug concentration. It may well be that such pharmacokinetic analysis will be useful in predicting toxicity, since the response of normal tissue to given drug concentrations should show less variability. With regard to antitumor effects, it is likely that additional approaches will be needed to define the relationship between drug concentration and therapeutic response, such as the integration of pharmacokinetics with biochemical pharmacology and cell kinetics.

## II. Routine Monitoring of Antineoplastic Drugs

The usefulness of drug level measurement in the routine clinical use of antineoplastic drugs depends on the identification during more complete pharmacokinetic studies of a single time point or small number of points reflecting the interindividual variation that may result in increased toxicity or a compromise of therapeutic effect. A few such applications have been demonstrated in the clinical setting, but most remain speculative and must be verified by future studies. Several characteristics of available chemotherapeutic agents make routine drug monitoring a desirable goal. The therapeutic index for most antitumor agents is extremely low. Thus, the likelihood of toxicity or a lack of therapeutic effect developing from fixed dosing is not to be ignored. Routine monitoring could be used to predict toxicity in these cases and so allow the physician to intervene prior to manifestation of clinical toxicity. In many instances, chemotherapeutic agents cause targettissue toxicity that cannot be detected at an early stage by routine clinical tests. Such is the case with adriamycin cardiotoxicity and bleomycin pulmonary toxicity. Definition of a pharmacokinetic parameter that would predict the toxicity sufficiently early to obviate these toxic effects would allow routine monitoring to be used in these cases. A number of factors may be responsible for interindividual variation in pharmacokinetics and the capricious nature of clinical toxicity, including wide variations in drug absorption, metabolism, or elimination. They may lead to considerable variability in clinical effectiveness of fixed doses. Routine monitoring may allow dose adjustment on a more rational basis than is presently used, i.e., nadir white blood count or platelet count. Renal or hepatic disease, which are often concomitant problems in patients with neoplastic disease, may significantly alter a drug's activity. Monitoring drug levels would allow the safe use

of chemotherapeutic agents in patients with such organ impairment, with adjustment of subsequent doses on a rational basis. The occurrence of saturation kinetics in drug uptake, transport, and elimination may result in severe toxicity with only small increments of drug dose. Although this has not been a recognized problem in the use of antineoplastic agents in the past, recent pharmacological studies of intraperitoneal 5-FU have suggested that saturation of an elimination process may play a role in the level at which toxicity occurs [66]. Drug interactions may significantly alter pharmacokinetics and clinical effects. With the common use of combination chemotherapy, such interactions should be systematically evaluated. Little work has been carried out as yet in this area. Finally, for drugs having a prolonged plasma halflife, patient compliance can be verified by blood level measurement. Certainly, the potential for acute toxicity such as nausea and vomiting may encourage patients to omit doses of oral medication.

While these reasons for monitoring are frequently appreciated in clinical practice, opportunities for the use of routine monitoring are limited by the lack of availability of simple, rapid, and yet dependable assay methods that are sufficiently inexpensive to allow their frequent use in patient care. However, recent years have witnessed a definite increase in reliable clinical pharmacokinetic studies suggesting a more specific role for routine drug level monitoring to improve either the safety or efficacy of antineoplastic therapy. The most thoroughly studied example is the use of methotrexate (MTX) in high-dose infusions. These infusions depend on rapid renal excretion of the drug for safe administration, and the ability of individual patients to excrete MTX is not always predictable on the basis of routine pretreatment screening tests of renal function. Since high-dose MTX infusions have been associated with severe myelosuppression, stomatitis, and a 6% incidence of drug-related fatalities [75], a clear need has been present to carefully monitor drug administration in an attempt to predict which patients are at increased risk for toxicity. This has been greatly simplified by the development of rapid, sensitive radioimmunoassays and competitive enzyme-binding assays for MTX in plasma, which make it possible to report results to clinicians within hours [2, 5, 12, 36, 46, 47, 54, 60]. The careful study of several high-dose MTX regimens has established the value of routine drug level monitoring in predicting toxicity [38, 56, 68, 76]. Stoller and co-workers found that patients treated with a 6-h infusion of 50-250 mg MTX/kg who had a plasma level of greater than 0.9  $\mu M$  at 48 h had a high likelihood of myelosuppression. This toxicity was not observed in patients with lower 48 h levels. They also determined that the administration of higher doses of leucovorin (100 mg/m<sup>2</sup>) effectively prevented this toxicity in some cases despite the continued presence of toxic levels of MTX [68]. Other studies have also suggested that such intensified rescue regimens may prevent toxicity among patients with delayed drug clearance [38]. The importance of adequate hydration and alkalinization of the urine has also been properly emphasized [56].

Although the recommendations of authors vary slightly as to the timing of MTX level determination for optimal clinical utility, monitoring practices for this drug are all based on similar principles. First, there is a critical threshold concentration of MTX for sensitive normal tissues, which must be exceeded before DNA synthesis is inhibited and tissue is damaged [16]. Second, the concentration or dose of leucovorin required to reverse MTX effect in vitro appears to increase with increasing MTX concentrations [59]. This work provides the rationale for escalation of leucovorin rescue in patients at risk for toxicity. Two groups have also shown that leucovorin dosage following high-dose MTX infusion can be minimized, and the rescue of malignant tissue theoretically limited, by basing leucovorin doses on measurement of serum MTX [40, 69]. Finally, there is a critical threshold for duration of exposure required to produce damage to sensitive tissues. Clinical toxicity seems to be correlated with both the duration of time that this threshold is exceeded and the degree of elevation of drug concentration above the threshold. In one study, infusions of up to 36 h resulted in minimal toxicity if adequate leucovorin was begun promptly, whereas longer infusions during which lower plasma MTX levels were maintained resulted in severe toxicity [33].

The measurement of MTX in cerebrospinal fluid (CSF) has become a necessary accompaniment of attempts to effectively treat CNS malignancy with this drug. The syndrome of acute MTX neurotoxicity has been associated with delayed clearance of the drug from the CSF following intrathecal injection [10]. Measurement of CSF levels of MTX may therefore be helpful clinically in distinguishing MTX neurotoxicity from malignant leptomeningitis. The demonstration of poor penetration of MTX into ventricular CSF following intralumbar injections of the drug [63] has provided one explanation for the high relapse rate of leukemic meningitis and has prompted the study of the Ommaya reservoir and other techniques of administration for patients with active CNS malignancy [I, II]. High-dose systemic MTX infusions have been shown to be effective in the treatment of CNS involvement with non-Hodgkin's lymphoma [64], and are currently under study as a mode of therapy for prophylaxis of leukemic and lymphomatous meningitis in high-risk patients. This type of therapy will require monitoring to document achievement of anticipated therapeutic MTX levels in the CSF and to aid in interpretation of the clinical outcome.

Although drug level monitoring has not been established for the routine clinical use of 5-fluoropyrimidines,

existing studies of the use of 5-FU illustrate both the possibilities and the difficulties with this approach. For example, the erratic plasma levels noted following orally administered 5-FU, with peak levels that are lower and delayed compared with the same dose given intravenously [14, 24, 32, 35, 52] has been used as an explanation for the lower response rates seen in some of the clinical trials in which these routes of administration are compared [4, 67]. The prolonged half-life of 5-FU when given with thymidine and the attendant host toxicity provides a second example of a rough correlation between plasma levels of drug and clinical toxicity or drug effect. This gives cause for optimism that the measurement of 5-fluoropyrimidine in plasma may be useful to clinicians.

There are several reasons why this has not occurred which are typical of the situation with most other antineoplastic agents as well. Published studies have been unable to quantitate 5-FU in plasma beyond 3 h [21, 24, 32], while the antimetabolic activity persists for a more prolonged period. It is known that 5-fluorodeoxyuridylate (5-FdUMP), the active metabolite that inhibits thymidylate synthetase, may persist in tissues for several days [18, 19, 55]. The cellular disappearance of 5-FdUMP, and perhaps ribonucleotide forms as well, determines the duration of drug effect and in part the degree of cytotoxicity. Differential sensitivity to 5-FU in two murine tumor lines has been correlated with the cellular clearance of this inhibitor [53]. Therefore, until plasma determination of 5fluoropyrimidine can be shown to have some relevance to the cellular determinants of drug effect, drug level monitoring will not reach its greatest utility. Finn and Sadee, using an isotope dilution mass spectrometric assay, observed a terminal half-life of 20 h in rats treated with 5-FU [32]. This value correlates more closely with the duration of 5-FU effect, and these authors suggested that this terminal half-life may reflect the elimination of 5-FUderived nucleotides from tissue. Unfortunately, the assay was not sufficiently sensitive to measure 5-FU in the terminal elimination phase from plasma of human subjects. The development and availability of analytical methods capable of measuring these low concentrations of 5-FU, 5-fluorouridine, and 5-fluorodeoxyuridine would allow this hypothesis to be fully tested and would possibly be very useful in identifying patients at risk for toxicity.

Two additional instances in which drug level monitoring may have a role have been suggested by recent work. Crooke and co-workers have demonstrated that the terminal elimination half-life for bleomycin in patients given an intravenous bolus increases exponentially as the creatinine clearance decreases below 25–35 ml/min [26]. Definite recommendations concerning the dosage modifications of bleomycin that may be necessary in patients with compromised renal failure will not be possible until the incidence and severity of drug toxicity are correlated with drug levels, plasma half-life, or renal function. These

studies are of great potential importance and should be forthcoming, since a sensitive and specific radioimmunoassay for bleomycin peptides is available [13, 70].

And finally, a provocative study by van Proojien and co-workers of the pharmacokinetics of ara-C in patients with acute myeloid leukemia (AML) has demonstrated a correlation between plasma half-life and remission induction in a small group of patients treated with 100 mg ara-C/m² every 12 h for 10–14 days [73]. Five patients with a second phase half-life of 6.6–10.7 min had a poor treatment response, whereas nine patients with a half-life of greater than 12.7 min had a complete bone marrow remission at 3 weeks. Since many patients with AML are treated by continuous infusion, this study leaves important questions unanswered, but it represents an extremely important attempt acquire pharmacokinetic data with relevance to the therapy of individual patients.

### Conclusions

Pharmacokinetics of antineoplastic agents can be considered with respect to comprehensive pharmacokinetic studies and routine drug monitoring. Ideally, the comprehensive studies should be completed in early investigative trials prior to the widespread clinical use of the drug to define the absorption, distribution, elimination, and metabolism of the drug. Only recently, with the development of new technological advances, has this become possible. Therefore, many of the commonly used antineoplastic agents are only now being clearly defined as to their pharmacokinetic behavior. To date, the clinical utility of monitoring has been defined only for MTX. Monitoring of other antineoplastic agents, such as 5-FU, melphalan, and bleomycin, is within the grasp of clinical pharmacology. Other agents, such as adriamycin and its analogs, await the development of facile but specific techniques and the definition of pharmacokinetic parameters that are of predictive importance. Although pharmacokinetics has much to offer in the area of cancer chemotherapy, the heterogeneous nature of the neoplastic process will probably require an integrative approach of pharmacokinetics, biochemical pharmacology, and cell kinetics to predict therapeusis or toxicity.

### References

- Abelson HT, Ensminger W, Rosowsky A, Uren J (1978) Comparative effects of citrovorum factor and carboxypeptidase G<sub>1</sub> on cerebrospinal fluid methotrexate pharmacokinetics. Cancer Treat Rep 62:1549
- Aherne GW, Piall EM, Marks V (1977) Development and application of a radioimmunoassay for methotrexate. Br J Cancer 36:608

- 3. Alberts DS, Chang SY, Chen HS, Evans TL, Moon TE (1979) Oral melphalan kinetics. Clin Pharmacol Ther 26:737
- Ansfield F, Klotz J, Nealon T, Ramirez G, Minton J, Hill G, Wilson W, Davis H Jr, Cornell G (1977) A phase III study comparing the clinical utility of four regimens of 5-fluorouracil. A preliminary report. Cancer 39:34
- Arons E, Rothenberg SP, daCosta M, Fischer C, Iqbal MP (1975)
   A direct ligand-binding radioassay for the measurement of methotrexate in tissues and biological fluids. Cancer Res 35:2033
- Banzet P, Jacquillat C, Civatte J, Puissant A, Maral J, Chastang, C, Israel L, Belaich S, Jourdain J, Weil M, Auclerc G (1978) Adjuvant chemotherapy in the management of primary malignant melanoma. Cancer 41:1240
- Benjamin RS, Wiernik PH, Bachur NR (1974) Adriamycin chemotherapy efficacy, safety, and pharmacologic basis of an intermittent single high-dose schedule. Cancer 33:19
- Bleyer WA (1978) The clinical pharmacology of methotrexate. Cancer 41:36
- Bleyer WA, Poplack DG (1978) Clinical studies on central nervous system pharmacology of methotrexate. In: Clinical pharmacology of anti-neoplastic drugs. Elsevier/North Holland Biomedical Press, Amsterdam, p 115
- Bleyer WA, Drake JC, Chabner BA (1973) Pharmacokinetics and neurotoxicity of intrathecal methotrexate therapy. N Engl J Med 289:770
- Bleyer WA, Poplack DG, Simon RM (1978) "Concentration x time" methotrexate via a subcutaneous reservoir: A less toxic regimen for intraventricular chemotherapy of central nervous system neoplasms. Blood 51:835
- Bohoun C, Duprey F, Boudene C (1974) Radioimmunoassay of methotrexate in biological fluids. Clin Chim Acta 57:263
- Broughton A, Strong JE (1976) Radioimmunoassay of bleomycin. Cancer Res 35:1418
- 14. Bruckner HW, Creasey WA (1974) The administration of 5-fluorouracil by mouth. Cancer 33:13
- 15. Buckpitt AR, Longo NS, Londer H, Boyd MR (1978) Assay of 5-fluorouracil and 5-fluorodeoxyuridine in plasma at the low nanogram level using high-pressure liquid chromatography. Proc Am Assoc Cancer Res 19:924
- Chabner BA, Young RC (1973) Threshold methotrexate concentration for in vivo inhibition of DNA synthesis in normal and tumorous target tissues. J Clin Invest 52:1804
- Chabner BA, Myers CE, Oliverio VT (1977) Clinical pharmacology of anticancer drugs. Semin Oncol 4:165
- Chadwick M, Chang C (1976) Comparative physiological disposition of 5-fluoro-2'-deoxyuridine and 5-fluorouracil in mice bearing solid L1210 lymphocyte leukemia. Cancer Treat Rep 60:845
- Chadwick M, Rogers I (1972) The physiological disposition of 5fluorouracil in mice bearing solid L1210 lymphocytic leukemia. Cancer Res 32:1045
- Chang SY, Alberts DS, Melnick LR, Walson PD, Salmon SE (1978) High-pressure liquid chromatographic analysis of melphalan in plasma. J Pharm Sci 67:679
- Clarkson B, O'Connor A, Winston L, Hutchison D (1965) The physiologic disposition of 5-fluorouracil and 5-fluoro-2'-deoxyuridine in man. Clin Pharmacol Ther 5:581
- Cohen JL, Brennan PB (1973) GLC assay for 5-fluorouracil in biological fluids. J Pharm Sci 52:572
- Cohen JL, Brown RE (1978) High-pressure liquid chromatographic analysis of 5-fluorouracil in plasma. J Chromatogr 151:237
- Cohen JL, Irwin LE, Marshall GJ, Darvey H, Bateman JR (1974)
   Clinical pharmacology of oral and intravenous 5-fluorouracil.
   Cancer Chemother Rep 58:723

- Colvin M (1978) A review of the pharmacology and clinical use of cyclophosphamide. In: Clinical pharmacology of antineoplastic drugs. Elsevier/North Holland Biochemical Press, Amsterdam, p 245
- 26. Crooke ST, Comis RL, Einhorn LH, Strong JE, Broughton A, Prestayko AW (1977) Effects of variations in renal function on the clinical pharmacology of bleomycin administered as an i.v. bolus. Cancer Treat Rep 61:1631
- 27. DeVita VT, Denham C, Davidson JD, Oliverio VT (1967) The physiological disposition of the carcinostatic 1,3-bis(2-chloroethyl)-1-nitrosourea (BCNU). Clin Pharmacol Ther 8:566
- D'Incalci M, Bolis G, Mangioni C, Morasca L, Garattini S (1978)
   Variable oral absorption of hexamethylmelamine in man. Cancer Treat Rep 62:2177
- D'Incalci M, Morazzoni P, Pantarotto C (1979) Gas chromatographic determination of hexamethylmelamine in mouse plasma. Anal Biochem 99:441
- Elson MK, Oken MM, Shafer RB, Broughton A, Strong J, Braun CT, Crooke ST (1978) Comparison of two radioimmunoassays and a microbiologic assay for bleomycin. Med Pediatr Oncol 5:213
- Fingl E, Woodbury DM (1975) General principles. In: The pharmacological basis of therapeutics, 5th edn. MacMillan, New York,
   p 1
- 32. Finn C, Sadee W (1974) Determination of 5-fluorouracil plasma levels in rats and man by isotope dilution-mass fragmentography. Cancer Chemother Rep 59:279
- Golde JH, Price LA, Harrap KR (1972) Methotrexate toxicity: Correlation with duration of administration, plasma levels, dose, and excretion pattern. Eur J Cancer 8:409
- Grochow LB, Colvin M (1979) Clinical pharmacokinetics of cyclophosphamide. Clin Pharmacokinetics 4:380
- 35. Hahn RG, Moertel CG, Schutt AJ, Bruckner HN (1975) A double-blind comparison of intensive course 5-fluorouracil by oral vs. intravenous route in the treatment of colorectal carcinoma. Cancer 35:1031
- 36. Hendel J, Sarek LJ, Vuidberg EF (1976) Rapid radioimmunoassay for methotrexate in biological fluids. Clin Chem 22:813
- Henderson ES, Adamson RH, Oliverio VT (1965) Metabolic fate of tritiated methotrexate. II. Absorption and excretion in man. Cancer Res 25:1018
- 38. Isacoff WH, Morrison PF, Aroesty J, Willis KL, Block JB, Lincoln TL (1977) Pharmacokinetics of high-dose methotrexate with citrovorum factor rescue. Cancer Treat Rep 61:1665
- Israel M, Pegg WJ, Wilkinson PM, Garnick MB (1978) Liquid chromatographic analysis of adriamycin and metabolites in biological fluids. J Liquid Chromatogr 1:795
- Jacobs SA, Santicky MJ (1978) Phase I trial of high-dose methotrexate with modified citrovorum factor rescue. Cancer Treat Rep 62:397
- Jones RB, Myers CE, Guarino AM, Dedrick RL, Hubbard SP, DeVita VT (1978) High volume intraperitoneal chemotherapy for ovarian cancer: Pharmacologic basis and early results. Cancer Chemother Pharmacol 1:161
- 42. Kirkwood JM, Frei E III (1978) 5-Fluorouracil with thymidine: A phase I study. Proc Am Assoc Cancer Res 19:159
- 43. Koch-Weser J (1975) The serum level approach to individualization of drug dosage. Eur J Clin Pharmacol 9:1
- 44. Kreis W, Gordon C, Gizoni C, Woodcock T (1977) Extraction and analytic procedures for cytosine arabinoside and 1-D-arabinofuranosyluracil and their 5'-mono-, di-, and triphosphates. Cancer Treat Rep 61:643
- 45. Leff P, Bardsley WG (1979) Pharmacokinetics of chlorambucil in ovarian carcinoma using a new HPLC assay. Biochem Pharmacol 28:1289

- Levine L, Powers E (1974) Radioimmunoassay for methotrexate.
   Res Commun Chem Pathol Pharmacol 9:543
- Loeffler LJ, Blum MR, Nelsen MA (1976) A radioimmunoassay for methotrexate and its comparison with spectrophotometric procedures. Cancer Res 36:3306
- 48. MacMillan WE, Wolberg WH, Welling PG (1978) Pharmacokinetics of fluorouracil in humans. Cancer Res 38:3479
- 49. Maddocks JL (1979) Assay of azathioprine, 6-mercaptopurine and a novel thiopurine metabolite in human plasma. Br J Clin Pharmacol 8:273
- Martin DS, Stolfi RL (1977) Thymidine enhancement of antitumor activity of 5-fluorouracil against advanced murine (CD8Fl) breast carcinoma (Abstract). Proc Am Assoc Cancer Res 18:126
- Martin DS, Stolfi RL, Spiegelman S (1978) Strikling augmentation of the in vivo anticancer activity of 5-fluorouracil by combination with pyrimidine nucleosides: An RNA effect (Abstract).
   Proc Am Assoc Cancer Res 19:221
- 52. Mukherjee KL, Curreri AR, Javid M, Heidelberger C (1963) Studies on fluorinated pyrimidines. XVII. Tissue distribution of 5fluorouracil-2-<sup>14</sup>C and 5-fluoro-2-deoxyuridine in cancer patients. Cancer Res 23:67
- Murinson DS, McMenamin M, Anderson T (1979) Clearance of acid soluble 5-fluorodeoxyuridine-5'-monophosphate in vivo as a determinant of 5-fluorouracil sensitivity (Abstract). Proc Am Assoc Cancer Res 20:251
- Myers CE, Lippman ME, Eliot HM, Chabner BA (1975a) Competitive protein binding assay for methotrexate. Proc Natl Acad Sci USA 72:3683
- Myers CE, Young RC, Chabner BA (1975b) Biochemical determinants of 5-fluorouracil response in vivo. J Clin Invest 56:1231
- 56. Nirenberg A, Mosende C, Mehta B, Gisolfi AL, Rosen G (1977) High-dose methotrexate with citrovorum rescue: Predictive value of serum methotrexate concentrations and corrective measures to avert toxicity. Cancer Treat Rep 61:779
- Peters JH, Murray JF (1979) Determination of adriamycin and aclacinomycin A in plasma by high-pressure liquid chromatography and spectrophotofluorometry. J Liquid Chromatogr 2:45
- Piall EM, Aherne GW, Marks VM (1979) A radioimmunoassay for cytosine arabinoside. Br J Cancer 40:548
- Pinedo HM, Zaharko DS, Bull JM, Chabner BA (1976) The reversal of methotrexate cytotoxicity to mouse bone marrow cells by leucovorin and nucleosides. Cancer Res 36:4418
- 60. Raso V, Schreiber R (1975) A rapid and specific radioimmunoassay for methotrexate. Cancer Res 5:1407
- Richens A Warrington S (1979) When should plasma drug levels be monitored? Drugs 17:488
- Rutty CJ, Connors TA (1977) In vitro studies with hexamethylmelamine. Biochem Pharmacol 26:2385

- 63. Shapiro WR, Young DF, Mehta BM (1975) Methotrexate distribution in cerebrospinal fluid after intravenous, ventricular, and lumbar injections. N Engl J Med 293:161
- 64. Skarin AT, Zuckerman KS, Pitman SW, Rosenthal DS, Moloney W, Frei E, Canellos GP (1977) High-dose methotrexate with folinic acid in the treatment of advanced non-Hodgkin's lymphoma including CNS involvement. Blood 50:1039
- Speyer JL, Collins JM, Dedrick RL, Brennan MF, Londer H, DeVita VT, Myers CE (1979) Phase I and pharmacological studies of intraperitoneal 5-fluorouracil. Proc Am Soc Clin Oncol 20:C-251
- Speyer JL, Collins JM, Dedrick RL, Brennan MF, Londer H, DeVita VT, Myers CE (1980) Phase I and pharmacologic studies of intraperitoneal 5-fluorouracil. Cancer Res (in press)
- 67. Stolinsky DC, Pugh RP, Bateman JR (1975) 5-Fluorouracil therapy for pancreatic carcinoma: Comparison of oral and intravenous routes. Cancer Chemother Rep 59:1031
- Stoller RG, Hande KR, Jacobs SA, Chabner BA (1977) Use of plasma pharmacokinetics to predict and prevent methotrexate toxicity. N Engl J Med 297:630
- 69. Stoller RG, Kaplan HG, Cummings FJ, Calabresi P (1979) A clinical and pharmacological study of high-dose methotrexate with minimal leucovorin rescue. Cancer Res 39:908
- 70. Strong JE, Broughton A, Crooke ST (1977) Specificity of antisera produced against bleomycin. Cancer Treat Rep 61:1509
- Sweat ML, Bryson MJ (1960) The role of phosphopyridine nucleotides in the metabolism of cortisol by peripheral tissues. Biochim Biophys Acta 44:217
- Tattersall MHN, Jarman M, Newlands ES, Holyhead L, Milstead RAV, Weinberg A (1978) Pharmaco-kinetics of melphalan following oral or intravenous administration in patients with malignant disease. Eur J Cancer 14:507
- van Proojien R, van der Kleijn E, Haanen C (1977) Pharmacokinetics of cytosine arabinoside in acute myeloid leukemia. Clin Pharmacol Ther 21:744
- Vogel S, Presant C, Ratikin G, Klahr C (1978) Phase I study of infusion 5-fluorouracil plus thymidine. Proc Am Assoc Cancer Res 19:232
- Von Hoff DD, Penta JS, Helman LJ, Slavik M (1977) Incidence of drug-related deaths secondary to high-dose methotrexate and citrovorum factor administration. Cancer Treat Rep 61:745
- Wang Y, Lantin E, Sutow WW (1976) Methotrexate in blood, urine, and cerebrospinal fluid of children receiving high doses by infusion. Clin Chem 22:1053
- Woodcock TM, Martin DS, Kemeny N, Young CW (1978) Phase I evaluation of thymidine plus fluorouracil in patients with advanced cancer. Proc Am Assoc Cancer Res 19:351

Received January 28, 1980